

Quality confirmation of RIKEN ^{186}Re using bifunctional chelating agents and derivatives

S. Oshikiri,^{*1,*2} M. K. Satake,^{*1,*2} H. Kato,^{*1,*2} Y. Komori,^{*1} K. Suzuki,^{*1,*2} T. Kobayashi,^{*2} A. Hino,^{*2} and H. Haba^{*1}

Rhenium-186 (half-life $T_{1/2} = 3.7186$ days) and ^{188}Re ($T_{1/2} = 17.003$ hours) emit beta rays appropriate for targeted radiotherapy use. These radioactive Re isotopes are used according to the tumor size and form radiotheranostic pairs with $^{99\text{m}}\text{Tc}$ for radiodiagnosis.¹⁾ However, ^{186}Re has not been investigated as well as ^{188}Re has.²⁾ One of the possible reasons is that it is difficult to obtain no-carrier-added ^{186}Re through the classical production method of the $^{185}\text{Re}(n, \gamma)^{186}\text{Re}$ reaction, while no-carrier-added ^{188}Re can be obtained using a $^{188}\text{W}/^{188}\text{Re}$ generator.^{1,3)} At RIKEN, we started to produce no-carrier-added ^{186}Re (RIKEN ^{186}Re) in the $^{186}\text{W}(d, 2n)^{186}\text{Re}$ reaction at the RIKEN AVF cyclotron. In this study, we selected DADT, ECD, MAG₃, and DMSA (Fig. 1) as model compounds, which had already been reported in many articles,^{4–7)} and evaluated radiolabeling efficiencies for these compounds to confirm the quality of RIKEN ^{186}Re , especially in terms of its usefulness as a radioisotope (RI) material for bifunctional chelating agents and derivatives.^{8,9)}

In this report, the method for DADT radiolabeling is described below as the representative among the model compounds.

- Step 1: RIKEN ^{186}Re (3.8 MBq) was dissolved in 0.05 M hydrochloric acid to prepare a ^{186}Re stock solution (138 MBq/mL). The radioactivity of ^{186}Re was determined using a germanium semiconductor detector and a dose calibrator.
- Step 2: 2.2 μL of the ^{186}Re stock solution in Step 1 was added to 63.8 μL of saline to prepare a ^{186}Re solution.
- Step 3: 3.0 μL of the ^{186}Re solution in Step 2 was mixed with 2.5 μL of DADT (1.0 μg), 1.4 μL of tin (II) dichloride dihydrate (10 μg), and 2.0 μL of L-tartaric acid (200 μg) aqueous solution.
- Step 4: The mixture in Step 3 was heated to 99°C and held for 15 min.
- Step 5: The radiolabeling yield of ^{186}Re -DADT was determined using the TLC method with a C18 reversed-phase TLC plate (NAGEL RP-18W/UV254) and eluted with acetone and 0.5 M ammonium acetate in a volume ratio of 13:7.

As a result, the radiolabeling yield of ^{186}Re -DADT at the specific radioactivity of 4.3 GBq/mmol was 86%. In Ref. 4), 88% of the radiolabeling yield of ^{188}Re -DADT at the specific radioactivity of 23 GBq/mmol was reported. To compare these results, each activity of ^{186}Re and ^{188}Re was converted to the amount of substance.

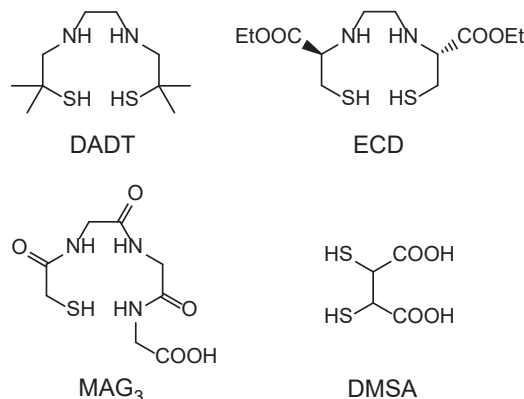


Fig. 1. Chemical structures of DADT, ECD, MAG₃, and DMSA.

Both 4.3 GBq of ^{186}Re and 23 GBq of ^{188}Re correspond to 3.3 nmol. This indicates that the ratio of the amount of DADT to that of $^{186}/^{188}\text{Re}$ was constant. Therefore, it was regarded that our result of labeling yield corresponded to that of ^{188}Re in Ref. 4). In a previous *in vivo* and *in vitro* evaluation, the radiolabeling of 222-MAMA(*N*-6-Ahx-OEt) with ^{186}Re was performed on 6.1 GBq/mmol.¹⁰⁾ It was suggested that RIKEN ^{186}Re is useful as an RI material for *in vivo* and *in vitro* studies.

Regarding the radiolabeling efficiencies for ECD, MAG₃, and DMSA, the radiolabeling yields were 51%, 46%, and 95%, respectively. It was revealed that these compounds are also able to form ^{186}Re complexes, although these are preliminary results.

In conclusion, our study revealed the availability of RIKEN ^{186}Re for radiolabeling and its feasibility as an RI material with bifunctional chelating agents and derivatives. By optimizing the labeling conditions in the future, RIKEN ^{186}Re is expected to be applied to targeted radiotherapy using bifunctional chelating agents, such as a peptide moiety in their structures.¹¹⁾

References

- 1) N. Lepareur *et al.*, *Front. Med.* **6**, 132 (2019).
- 2) L. Uccelli *et al.*, *Molecules* **24**, 640 (2019).
- 3) G. Makris *et al.*, *Mol. Imaging Biol.* **23**, 52 (2021).
- 4) J. M. Jeong *et al.*, *Nucl. Med. Biol.* **28**, 197 (2001).
- 5) T. -Y. LuO *et al.*, *Nucl. Med. Biol.* **31**, 671 (2004).
- 6) G. W. Visser *et al.*, *J. Nucl. Med.* **34**, 1953 (1993).
- 7) K. Kothari *et al.*, *Appl. Rad. Iso.* **51**, 43 (1999).
- 8) G. Liu *et al.*, *Anticancer Agents Med. Chem.* **7**, 367 (2007).
- 9) U. Choudhry *et al.*, *Dalton Trans.* **3**, 311 (2003).
- 10) D. W. Demoin *et al.*, *Nucl. Med. Biol.* **43**, 802 (2016).
- 11) V. A. Sanders *et al.*, *Nucl. Med. Biol.* **68–69**, 1 (2019).

*1 RIKEN Nishina Center

*2 RI Research Department, FUJIFILM Toyama Chemical Co., Ltd.